

Figure 3, cont'd.

---

- (d) About 1/2 of the distance inward;
- (e) About 3/4 of the distance inward;
- (f) At the center of the affected area.

For smaller kills, collect the 1 fresh and 1 preserved sample at locations (a),(b),(d),(f); that is, samples should be collected outside the kill area, on the outer fringe of it, about halfway into it, and then at the center or worst area of the kill (for a total of 8 samples).

4. We would like to help sample, especially in moderate / large kills, so we'd really appreciate your contacting us as soon as you can when you learn that a substantial kill is occurring (or has just occurred), so that we can mobilize quickly and get to the site to collect many other types of samples that would be helpful in tracking this dinoflagellate.
5. To facilitate testing for potential toxic dinoflagellate activity, it would be best to receive the samples as quickly as possible. Please send them to us by State courier mail (# 536121), or call us so that we can make other arrangements. Include a note that briefly describes the kill (date and time, types of fish affected, how dying fish looked or were acting, kill location, whether birds are eating the dead/dying fish, and other details you notice that might be of interest). Also, mark the bottles so that we can determine where, within the kill area, each sample was collected.

We will be able to confirm the presence of this toxic dinoflagellate within 1 day after receiving water samples (the procedure involves settling the preserved material overnight). However, it often requires several days to 2-4 weeks to confirm toxic activity, depending on whether the alga has encysted by the time we receive the live (fresh, unpreserved) samples.

We're grateful for your help.

---